# Unraveling Microbiome: The Role of Microbiota in Patients' Response to Oncological Treatment and Its Influence on Host-biochemistry

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Despite the implication of viruses and bacteria in oncogenesis, commensal microbes play a beneficial role in anticancer therapy. Microbiota regulates responses made to various types of cancer chemotherapy by affecting the mechanism of action and toxicity of these therapies. From birth, microbiota confers innate and acquired immune responses and its role in modulating health and diseases has recently been appreciated. Increasing evidence also suggests that microbiota affects the pattern of body energy balance thus; they have a correlation with obesity, obesityrelated complications and insulin resistance. Hence, microbiota has attracted the interest of researchers. However, there is little, if any, robust evidence-based review on their role in patients' response to cancer therapy and their influence on host-biochemistry thus, this review aimed to achieve that.

Keywords: Biochemical parameters, cancer chemotherapy, commensal microbes, microbiome, microbiota.

### INTRODUCTION

although, Microbiome, frequently interchangeably with microbiota, are not the same. Microbiota, since its coining in 2001, is used to describe the ecological community of microorganisms (symbiotic, commensal, pathogenic) whiles the microbiome encompasses genomes and all products of microbiota within the body (Orlandi et al., 2019). In other words, the microbiome is a more holistic, complex and diverse ecosystem pertaining to all host-associated microorganisms inhabiting epithelial barriers throughout the body and this includes bacterial microbiome (bacteria), archael microbiome (archaeal), virome (eukaryotic virusesand bacteriophages), mycobiome (fungi) as well as meiofauna (helminthic worms and unicellular protozoa) (Zitvogel et al., 2018). The microbiome is acquired via birth by means of vertical transmission (Zitvogel et al., 2018) and stays exposed to change by environmental factors such as nutrition throughout life. Also, microbiota influence brain and body development (Smith, 2015)as organisms raised in a controlled environment, that prevents the growth of microbiota (germ-free organisms) have poorly developed physiological conditions such as innate immunity (Raza et al., 2018). Mainly, it ensures homeostasis in the body especially, in the oral cavity, gastrointestinal tract, vagina and skin and this has a correlation with health status. A disruption in the microbial balance causes "dysbiosis" which subsequently leads to pathological conditions such as a chronic inflammatory disorder (Zitvogel et al., 2018). Following the tremendous discovery made by Williams B. Coley where anti-cancer responses were stimulated in cancer patients injected with heat-inactivated Streptococcal microorganisms, anticancer immunotherapy, in other words, the role of the system as a modulator of tumor proliferation/growth gained the attention of researchers (Bashiardes et al., 2017). According to Zitvogel et al. (2018), from birth, microbiota confers innate and acquired immune responses and its role in modulating health and diseases has recently been elucidated.

Cancer has been described as the second leading cause of death worldwide and it occurs as a result of intracellular accumulation of spontaneous mutations during DNA replication, combined with other environmental factors and lifestyle habits over a long period of time (Tomasetti Vogelstein, 2015; Ashford et al., Environmental factors such as ultraviolet radiation, toxic substances and infectious agents among other factors have been termed to be cancer predisposing factors (Anand et al., 2008). Following tobacco, infections maybe the next important preventable cause of cancers and the interest in infectious causes of cancers has ebbed and flowed (Kuper et al., 2000a). The microbiome has been reported to be involved in the initiation and exacerbation of various forms of cancers at epithelial barriers (Zitvogel et al., 2018) and in sterile environments (Dzutsev et al., 2015). **Emphasizing** involvement of the suitable microenvironments in carcinogenesis is the demonstrated carcinogenic pathway of the first discovered oncovirus, Rous sarcoma virus which induced in adult birds cancer at sites of injection or injury and could not induce cancer in sterile embryos. In addition to this, one-quarter of the skin cells contained in aged and sun-exposed eyelids having clonally expressed carcinogenic mutants similar to those in squamous cell carcinoma but, maintain normal dermatological physiology instigating without

carcinogenic effects (Dzutsev et al., 2017). Moreso, epidemiological studies based on the analysis of oral, fecaland tissue samples to ascertain the role of microbiota and dysbiosis in carcinogenesis reported the only bacterium recognized by the International Agency for Research on Cancer (IARC) as a group I human carcinogen, *Helicobacter pylori (H. pylori)* as the underlying etiology of stomach cancer (Dzutsev et al., 2017). Contrary to this, the anti-cancer response has been stimulated in superficial bladder cancer patients with intravascular injections of *Mycobacterium bovis (M. bovis)* (Bashiardes et al., 2017), therefore indicating microbiome triggered immune-anticancer response. In fact, the microbiome can either be tumor-suppressive or oncogenic (Fullbright et al., 2013; Gagnaire, 2017).

Despite the complexity and multi-factorial nature of carcinogenesis (cancer onset), there seems to be a lot of evidence on microbial carcinogenesis. Enhanced capacity of amino acid metabolism, lipid metabolism and digestive system has been observed in a treatment group having the highest population of beneficial bacteria (Liu, 2018). It implies that microbiotas also have an essential impact on the biochemistry of a host.

In healthy individuals, the abundance of various microbiota varies greatly in accordance with the host's immune system, environmental conditions, infections and use of antibiotics, as well as genetics and diet (Wirth et al., 2020) which the latter (diet) has emerged as an essential influencer of microbiota composition and function (Zmora et al., 2019). Also, mouse models have shown deterioration of gut microbiota by perturbing forces such as antibiotics and industrialized diets (Sonnenburg and Sonnenburg, 2014; Korpela, 2016; Schulfer et al., 2018). In addition to this, genetically similar populations have shown different microbiotas due to lifestyle differences such as farming practices and water sources (Fragiadakis et al., 2019; Jha et al., 2018; Gomez et al., 2016; Morton et al., 2015).

However, the distal gut microbiota of an adult human is typically dominated by two bacterial phyla; the Firmicutes and Bacteroidetes (Mahowald et al., 2009) and the relative variation in the abundance of these two bacterial phyla seems to be related to obesity. A relative greater abundance of Firmicutes has been reported in obese-leptin deficient mice as compared to the microbiota of lean-control mice. Similarly, when germ-free mice were colonized with the microbiota (Firmicutes) from an obese donor, they manifested an increase in body fat despite having the same feed intake. Obese-patients having much abundance of Firmicutes and significantly fewer abundance of Bacteroidetes compared with lean-controls, when subjected to the same diet as the controls, subsequently had a relative increase in Bacteriodetes which positively correlated with weight loss (Indiani et al., 2018). However, there is little, if any, robust evidencebased review on their role in patients' response to cancer therapy and their influence on host-biochemistry thus, this review aimed to achieve that.

## MICROBIAL CARCINOGENESIS

Prior to the end of the 19<sup>th</sup> century, the discoveries made in medical microbiology proved the involvement of bacteria and their etiology for major diseases and this pragmatically contributed to improved treatment. Not surprisingly, this perhaps led to the birth of the idea that bacterial infections might be the cause of carcinogenesis. However, these observations did not take into

consideration the long-time lag between the initiation of carcinogenesis and onset of the overt disease. Just as the bacterial infection would not be thought of as the cause of cystic fibrosis in patients, the presence of bacteria at sites of carcinomas does not necessarily mean they are the cause (Lax and Thomas, 2002). Notwithstanding that, cell transformation could have been caused bacterium/bacteria long before the manifestation of cancer and bacterium/bacteria could have been cleared from the body long before its full effects were seen. Lax and Thomas (2002), further stated that the involvement of bacteria in carcinogenesis remains partly controversial since there is no clear evidence on the molecular mechanism(s) by which bacteria cause carcinogenesis. Aside from that, the recognition of *H. pylori*as the cause of gastric cancer is based on epidemiological studies thus, its role in the emergence of stomach cancer is not straightforward (Falk et al., 2000). The diverse genetic nature of humans further complicates it as humans exhibit different susceptibilities to *H. pylori*infection and stomach cancer development (Magnusson et al., 2001). Furthermore, animal models have been used to provide more convincing evidence of other diseases. However, animal models used to demonstrate H. pyloriinfection resulted in a pathological display different from that seen in the human infection (Lax and Thomas, 2002, Dağ et al., 2016). Contrary, a disruption of the repertoire of gut microbiota, which is often known as "dysbiosis", is linked to several pathological conditions, including cancer (Carding et al., 2015) and they may elicit carcinogenesis by causing immune evasion or inflammatory disorders, DNA damage or direct tissue damage, in the case of oncovirus infection (Curty et al., 2019) (shown in Figure 1.). Models of bacteria-mediated carcinogenesis of several microbial species including, Enterococcus faecalis (E. faecalis), Streptococcus gallolyticus (S. gallolyticus), enteropathogenic Escherichia coli (E. coli), Salmonella enterica (S. enterica), H. hepaticus and Fusobacterium nucleatum (F. nucleatum) have also been characterized in mice (Kostic et al., 2013; Sears and Garrett, 2014).F. nucleatum has been etiologically linked to carcinogenesis of sporadic colorectal cancer (CRC) and E. coli strains to harbor a genomic virulence island have been reported to be capable of causing DNA damage and chromosomal instability in the host (Wirth et al., 2020). Besides, archae have also been found in fecal samples of colorectal cancer patients (Mira-Pascual et al., 2014). Viruses such as Hepatitis B virus (HBV), Epstein-Barr virus (EBV) and Human papilloma virus (HPV) have been accepted as carcinogenic owing to their direct mechanistic single geneinduced cell transformation effects (Kuper et al., 2000a). The chronic form of HBV infection is estimated to be the definite cause of hepatocellular carcinoma based a geographical correlation, cohort and case-control studies, as well as clinical and laboratory investigations (Kuper et al., 2000b). It is responsible for more than half of all liver cancer cases globally (Kuper et al., 2000a). Approximately, 2 billion people have been infected with HBV and about 350 million people are chronic carriers of the virus with the highest prevalence being in Asia and Africa. It can be transmitted vertically (from mother to infant) or sexually (Kuper et al., 2000a).

In HBV infection, the HBV-DNA may be integrated into the host genome which potentially results in insertional mutagenesis involving structural changes such as deletion of host DNA; translocation, duplication of HBV-DNA and amplification of host DNA (Robinson,

1994). The x protein coded for by the HBV-genome is responsible for the inactivation of *P53* and other tumor suppressor genes. It has been reported that, x protein may pay its carcinogenic quota via the transcription of methyl transferases, thus causing regional hyper-methylation of DNA which causes the silencing of tumor suppressor genes or may cause general hypo-methylation resulting in chromosomal instability. Hence, it plays a vital role in

hepatocarcinogenesis. Aside from the anti-apoptotic effect of *x* protein, it also instigates a pro-apoptotic effect. These contradicting effects are yet to be explained (Kew, 2011). Despite the strenuous and pragmatic efforts made to explain the molecular effects of HBV genome, direct viral effects of HBV are unlikely to be the main causes of liver carcinogenesis since not all hepatocellular carcinoma tumors contain HBV integrands (Kuper et al., 2000a).

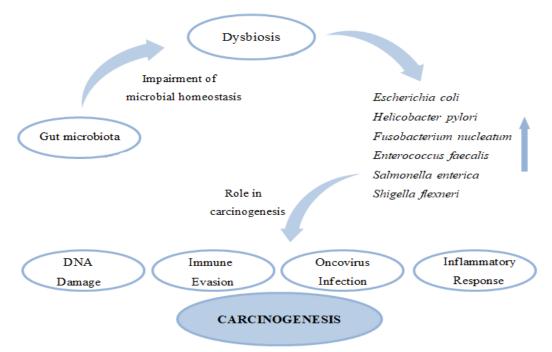


Figure 1. Illustrates dysbiosis-related carcinogenesis. Bacteria during gut dysbiosis can secrete toxins that may interfere with host cell growth, finally, predisposing the host organism to cancer development. Microbiota may elicit their carcinogenetic effect by either evading host-immune system or by triggering an inflammatory response, DNA damage or by directly instigating tissue damage (oncoviruses)

The member of the Herpes family of viruses, EBVwhich is made up of linear DNA, after infection, gains a circular shape to form EBV episomes with membranous envelope and glycoprotein spikes. Its genome codes for Epstein-Barr Nuclear Antigen 1 (EBNA1), EBNA2 and lymphocyte membraneassociated protein (LMP1) which may interfere with P53 mediated apoptosis. The z protein which triggers the switch of EBV from latent to lytic infection can interact directly with tumor suppressor protein, P53in vitro and in vivo. EBV is an established carcinogen, having conclusive evidence with respect to non-Hodgkin's lymphoma and nasopharyngeal carcinoma (IARC, 1997). EBV has been associated with other forms of cancer including gastric carcinoma, but evidence surrounding these claims are weak (Kuper et al., 2000a).

Human papillomaviruses are sexually transmitted viruses and have high tissue-specificity and mainly infect the basal cells of the squamous epithelium in the genital tract, skin and upper respiratory tract. It has been associated with vulva, anus, penis, head and neck cancers but, it is emphatically known for its cervical cancer etiology (Vaccarella et al., 2006). Furthermore, a case-control study conducted by IARC has labeled HPV strains 16 and 18 as definite human carcinogens, and types 35, 45, 51, 52, 58 and 59 have been referred to be possibly related to carcinogenesis (Pisani et al., 1997). The carcinogenic

effect of HPV is due to its ability to integrate into the host genome and code for the genes E6 and E7. These two genes are important for carcinogenesis and have been demonstrated to be capable of immortalizing the primary human genital keratinocytes *in vitro* (Pao et al., 1996). According to Pao et al. (1996), its ability to code for the E6 and E7 genes emanates from the disruption of the E2 gene during integration into the host genome. In other words, E2 gene has an anti-carcinogenic effect.

# RELEVANCE OF MICROBIOTA IN CANCER THERAPY

Despite the implication of viruses and bacteria in oncogenesis, commensal microbes play a beneficial role in anticancer therapy (Perez-Chanona and Trinchieri, 2016). Anti-cancer treatments may evoke a tumor-destructive immune response by altering the microenvironment (Shirota et al., 2012) and in a study conducted, subcutaneous tumors failed to respond to immunotherapy and platinum chemotherapy after antibiotics treatment, which is a microbiota perturbing force (Iida et al., 2013). This suggests that microbiome plays a crucial role in the efficacy of a number of anticancer therapeutic approaches (Sivan et al., 2015; Iida et al., 2013; Viaud et al., 2013). Microbiota regulates the responses made to various types of cancer chemotherapy by affecting the mechanism of action and toxicity of these therapies. In view of this,

interventions on microbiome may be pivotal to improving toxicity that may emerge from anti-cancer therapy as well as improving the efficacy of anti-cancer therapies (Nayak and Turnbaugh, 2016; Fessler and Gajewski, 2017). This can be substantiated using the crosstalk between gut microbiota and myeloid cells for the production of reactive oxygen species (ROS) in the case of oxaliplatin and its induction of an anticancer T cell response in the case of cyclophosphamide (Roy and Trinchieri, 2017).

In the study conducted by Iida et al. (2013), the tumorinfiltrating myeloid-derived cells of antibiotics-treated or germ-free mice resulted in lower production of cytokine and tumor necrosis after CpG oligonucleotide treatment and deficient production of ROSand cytotoxicity after chemotherapy, thus, responded poorly to cancer therapy. In this light, an intact commensal community (microbiota) is vital to the success of cancer therapy. Viaud et al. (2013) also demonstrated the relevance of microbiota in response to cancer therapy using tumor-bearing germ-free mice (mice treated with antibiotics to kill Gram-positive bacteria). According to the study, the mice had reduced "pathogenic" T helper 17 (pT<sub>H</sub> 17) and their tumors were resistant to cyclophosphamide. However, after an adoptive transfer of pT<sub>H</sub> 17 cells, the antitumor efficacy of cyclophosphamide was restored. Consequently, Grampositive bacteria shape the efficacy of cyclophosphamide since they stimulate the generation of pT<sub>H</sub> 17 cells as well as memory T<sub>H</sub> 1 immune response (Viaud et al., 2013). Oral administration of Bifidobacterium alone yielded a tumor control at a degree same as that of programmed cell death protein 1 ligand (PD-L1)-specific antibody therapy, and when administered concomitantly with PD-L1, they nearly abolished tumor outgrowth (Sivan et al., 2015). Lactobacillus rhamnosus GG (LGG) is a gut bacterium having anti-inflammatory effects (Khailova et al., 2017; Wang et al., 2017; Fong et al., 2016) and has been studied mostly as a probiotic due to its anti-inflammatory effect (Lee et al., 2014). LGG demonstrated a preservative effect on gut microbiota balance and on intestinal epithelial barrier functionality in radiation-mediated gut epithelial injury in animal models (Chang et al., 2018; Riehl et al., 2018; Zhang et al., 2017) and its administration in cancer patients has been pictured to have several potential benefits (Banna et al., 2017). Additionally, several in-vitro studies in tumor models including, colorectal, ovary and breast cancer tumor models have proven that, LGG has an anti-metastatic effect (Orlando et al., 2016; Nouri et al., 2016; Zhao et al., 2017). Also, treatment with LGG reduced tumor mass by modulating gut commensal microbiome in rat dimethyl hydrazine-induced colon cancer (Cai et al., 2016). Moreover, oral administration with Lactobacillus casei reduced the recurrence of superficial bladder cancer (Aso and Akazan, 1992). Also, according to Soyucok et al. (2020), the lactic acid bacterium Lactobacillus helveticus plays an essential role in the hydrolyses of milk proteins into bioactive peptides. Soyucok et al. (2020) further stated that, L. helveticus contributes essentially to health promotion as it modulates immune responses and intestinal microbiota by preventing gastrointestinal infections.

Furthermore, an evaluation of bacterial composition conducted on animals treated with CpG-ODN/a-IL-10 ab reported different bacterial species that correlated with the immune response against the tumor. For instance, the presence of the gram-negative *Alistipes genera* in the feces positively correlated with TNF production in the tumor (Farrokhi, 2019). In view of this, it can be concluded that

Alistipes genera provided an anticancer response by contributing to an increase in the number of TNFproducing myeloid cells in the tumor tissue. Additionally, Microbiotas may provide a positive response to cancer therapy through the production of SCFAs. SCFAs such as butyrate and propionate have been demonstrated to have an anti-cancer effect in CRC and lymphoma by inhibiting host's tumor cells histone deacetylases (HDACs) (Wei et al., 2016; Jan et al., 2002), which are responsible for regulating the expression and activity of proteins involved in the initiation and progression of cancer (Glozak and Seto, 2007). The interaction between bacterial microbiota and anticancer chemotherapeutics can affect the efficiency of these anticancer chemotherapeutics. For instance, the activity of 10 out 30 chemotherapeutics has been reported to have been suppressed in the presence of nonpathogenic bacterial species E. coli and Listeria welshimeri (Lehouritis et al., 2015). This further suggests that microbiota modulates cancer therapy.

It is becoming evident that, aside from the impact of gut microbiotas on cancer therapy, cancer therapy may in turn affect gut microbiotas. Chemotherapy may affect metabolic pathways by causing profound dysbiosis (Alexander et al., 2017; Montassier et al., 2015). In the course of chemotherapy, antibiotics are frequently administered to patients and this should be given much attention as it has been proven that; concurrent administration of antibiotics negatively affects the outcomes of cancer immunotherapy (Derosa et al., 2018). The administration of microbial consortia (probiotics) in an attempt to improve human health is nothing new, as efforts are still in progress to modulate the gut microbiota since the early 1900s when Metchnikof theorized that, administration of microbes could have beneficial effects (Podolsky, 2012). However, some preclinical and clinical studies conducted on commercial probiotics reported varied results. Although some reported positive potentials of the probiotics (Zhu et al., 2011; Appleyard et al., 2011), other studies with the same probiotics reported deleterious effects of the probiotics, with increased tumor penetrance and multiplicity (Arthur et al., 2013). Theoretically, the timing of probiotic administration, among other factors was explained to have contributed to the differences in effect. Clinical trials to study the impact of probiotics administration have been done in cancer patients but most of these studies focused on their impact on changes in microbiota composition and not specifically ascertain their influence on outcomes of cancer therapies (Helmink et al., 2019). Such report includes the administration of probiotics to patients with CRC which resulted in an alteration in the gut microbiota by increasing the abundance of butyrate-producing microbes in mucosal and fecal samples following administration (Hibberd et al., 2017). This insinuates that probiotics still have a promising effect of restoring microbiota composition or correcting dysbiosis. However, studies involving the impact of commercially available probiotics on microbiota should not be neglected as the content of commercially available probiotics may vary significantly from what is advertised (Kolaček et al., 2017; Morovic et al., 2016).

# EFFECTS OF MICROBIOTA ON BIOCHEMICAL PARAMETERS

Increasing evidence suggests that gut microbiota may influence weight-gain via several inter-dependent pathways such as short-chain fatty acids signaling, modification of behavior, controlling of appetite,

modulating of inflammatory responses within-host (Bliss and Whiteside, 2018) and energy harvesting (Ley et al., 2006). Also, dysbiosis may also lead to altered host inflammation status as well as liver disease (Boulangé et al., 2016; Minemura and Shimizu, 2015) as indicated in the summary(shown in Figure 2). In terms of energy, a balance should exist between energy intake and expenditure. An imbalance where the former (intake) exceeds the latter (expenditure) will result in obesity, which is a complex and global epidemic disease (Bliss and Whiteside, 2018), obesity-associated complications and

insulin resistance (Shen et al., 2013; Prieto et al., 2018). Proteobacteria, are Gram-negative bacteria with lipopolysaccharides (LPS) in their outer membranes. They have been reported by Prieto et al. (2018) to have a correlation with ghrelin levels. Chang et al. (2003) reported that LPS directly stimulated the gastric mucosa to synthesize and secret ghrelin in rats, which is considered as a therapeutic effect against the endotoxic shock produced by LPS. In this context, proteobacteria or LPS-containing bacteria can be termed to have a positive correlation with ghrelin.

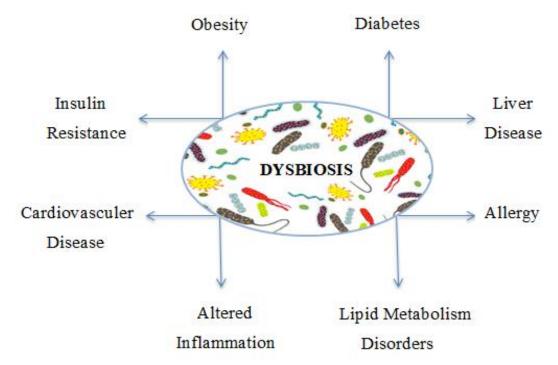


Figure 2. Dysbiosis may cause many diseases by changing intestinal permeability and the biochemical profile of the host.

Prieto et al. (2018) further stated that Desulfovibrio and Alistipes indictintus, which are LPS-containing bacteria also had a correlation with plasmatic insulin levels and hence they are involved in glucose intolerance. LPSinduced metabolic endotoxemia has been reported to be the first step in the development of insulin resistance diabetes (Type II diabetes) and this has been demonstrated with an experimental infusion of LPS which resulted in hyperglycemia and hyperinsulinemia (Cani et al., 2007). Leptin is a hormone secreted by fat cells to inhibit feeding by eliciting its effect via the hypothalamic pathway in order to achieve energy balance and body weight regulation (Jéquier, 2002). Gut microbiota has been reported to have crosstalk with this hormone since it is capable reducing hypothalamic sensitivity to leptin by leptin resistance-associated suppressor cytokine signaling 3 (Schéle, 2013). Medina-Vera et al. (2019) reported that, type 2 diabetic patients exhibiting intestinal dysbiosis characterized by an increase in Prevotella copri, with a dietary intervention independently of antidiabetic drugs had decreased Prevotella copri with increased Faecalibacterium prausnitzii and Akkermansia muciniphila (two bacteria known for their anti-inflammatory effects). Additionally, patients also

exhibited a significant decrease in glucose, total and low-density lipoprotein (LDL) cholesterol, free fatty acids (FFAs), triglycerides, glycosylated hemoglobin (HbA1c) and an increase in antioxidant activity. This further elaborates the biochemical parameters modulating the effect of the microbiota.

In addition, certain commensal bacteria are known to produce essential micronutrients such as vitamin K and other components of vitamin B. Some members of the family Bacteroides have been reported to be capable of synthesizing the anti-diabetic linoleic acid, catabolizing secondary bile acids and capable of breaking down phenolic compounds. Furthermore, some gut microbiotas are capable of modifying some amino acids by means of decarboxylation into signaling molecules such as; histidine into histamine and glutamate into gammaaminobutyric acid (GABA) (Mohajeri et al., 2018). Additionally, the commensals of the gut can also produce hormone-like metabolites known as short-chain fatty acids (SCFAs) by fermenting dietary fibers in the large intestine. These SCFAs when produced, they are transported via the bloodstream to the liver, where they are used as a source of energy. SCFAs also play an essential role in controlling the metabolism of glucose and lipid by affecting the secretion of a peptide hormone, peptide YY (PYY) and glucagon-like peptide 1 (GLP-1) (Clarke et al., 2014).

### CONCLUSION AND RECOMMENDATION

Microbiota is an essential integral human component that keeps evolving with time. Its role in response to cancer therapies or efficacy of cancer therapies is inevitable. Also, an intact microbiota is important for the maintenance of a healthy host-biochemistry. Although viral microbiotas such as HBV, EBV and HPV have been reported as carcinogenic, there are still discrepancies on the carcinogenicity of bacteria microbiome. Subsequent studies should be conducted on the influence of microbiota on response to cancer therapy and restoration of gut microbiota after treatment. Studies should be conducted on the molecular mechanism(s) of bacteria carcinogenicity. Further exploration should be done on the anticarcinogenic effects of the x protein coded for by the HBV genome and the E2 gene which is also coded for by the HPV genome. Further studies should also be conducted on the correlation between microbiota and biochemical parameters.

#### **Conflict of Interest**

The authors declared that there is no conflict of interest.

### REFERENCES

Alexander JL, Wilson ID, Teare J, Marchesi JR, Nicholson JK, Kinross JM. 2017. Gut microbiota modulation of chemotherapy efficacy and toxicity. Nature Reviews Gastroenterology Hepatology. 14: 356–365.

Anand P, Kunnumakkara AB, Sundaram C, Harikumar KB, Tharakan ST, Lai OS, Sung B, Aggarwal BB. 2008. Cancer is a preventable disease that requires major lifestyle changes. Pharmaceutical Research. 25: 2097–2116.

Appleyard CB, Cruz ML, Isidro AA, Arthur JC, Jobin C, Simone CD. 2011.Pretreatment with the probiotic VSL#3 delays transition from infammation to dysplasia in a rat model of colitisassociated cancer. American Journal of Physiology-Gastrointestinal and Liver Physiology. 301: 1004–1013.

Arthur JC, Gharaibeh RZ, Uronis JM, Perez-Chanona E, Sha W, Tomkovich S, Mühlbauer M, Fodor AA, Jobin C. 2013. VSL#3 probiotic modifes mucosal microbial composition but does not reduce colitis-associated colorectal cancer. Scientific Reports. 3: 2868.

Ashford NA, Bauman P, Brown HS, Clapp RW, Finkel AM, Gee D, Hattis DB, Martuzzi M, Sasco AJ, Sass JB. 2015. Cancer risk: Role of environment. Science 347: (6223),727.

Aso Y, Akazan H. 1992. Prophylactic effect of a Lactobacillus casei preparation on the recurrence of superficial bladder cancer. BLP Study Group. Urologia Internationalis. 49: 125–129.

Banna GL, Torino F, Marletta F, Santagati M, Salemi R, Cannarozzo E, Falzone L, Ferrau F, Libra M. 2017. Lactobacillus rhamnosus GG: An Overview to Explore the Rationale of Its Use in Cancer. Frontiers Pharmacology. 8: 603.

Bliss ES, Whiteside E. 2018. The Gut-Brain axis, The Human Gut microbiota and their Integration in Development of Obesity. Front Physiology. 9: 900.

Boulangé CL, Neves AL, Chilloux J, Nicholson JK, Dumas ME. 2016. Impact of the gut microbiota on inflammation, obesity, and metabolic disease. Genome Medicine. 8:42.

Cai S, Kandasamy M, Rahmat JN, Tham SM, Bay BH, Lee YK, Mahendran R.2016.

Lactobacillus rhamnosus GG Activation of Dendritic Cells and Neutrophils Depends on the Dose and Time of Exposure. Journal of Immunology Research.7402760: 8.

Cani PD, Amar J, Iglesias MA, Poggi M, Knauf C, Bastelica D, Neyrinck AM, Fava F, Tuohy KM, Chabo C, Waget A, Delmee E, Cousin B, Sulpice T, Chamontin B, Ferrieres J, Tanti JF, Gibson GR, Casteilla L, Delzenne NM, Alessi MC, Burcelin R. 2007. Metabolic endotoxemia initiates obesity and insulin resistance. Diabetes; 56(7):1761–72.

Carding S, Verbeke K, Vipond DT, Corfe BM, Owen LJ. 2015. Dysbiosis of the gut microbiota in disease. Microbial Ecology Health and Disease 26: 26191.

Chang CW, Liu CY, Lee HC, Huang YH, Li LH, Chiau JS, Wang TE, Chu CH, Shih SC, Tsai TH, Chen YJ. 2018. Lactobacillus casei variety rhamnosus Probiotic Preventively Attenuates 5-Fluorouracil/Oxaliplatin-Induced Intestinal Injury in a Syngeneic Colorectal Cancer Model Front Microbiology. 9: 983.

Chang L, Zhao J, Yang J, Zhang Z, Tang C. 2003. Therapeutic effects of ghrelin on endotoxic shock in rats. European Journal of Pharmacology. 473:171–6.

Clarke G, Stilling RM, Kennedy PJ, Stanton C, Cryan JF, Dinan TG. 2014. Gut microbiota: The neglected endocrine organ. Molecular Endocrinology. 28: 1221–1238

Curty G, De Carvalho PS, Soares MA. 2019. The Role of the Cervicovaginal Microbiome on the Genesis and as a Biomarker of Premalignant Cervical Intraepithelial Neoplasia and Invasive Cervical Cancer. International Journal

Dağ S, Sözmen M, Cihan M, Tunca R, Kurt B, Devrim AK, Özen H. 2016. Gastric Helicobakter-like organisms in stray cats: identification, prevalence and pathologic association. Pakistan Veterinary Journal. 36(2): 199-203.

Derosa L, Hellmann MD, Spaziano M, Halpenny D, Fidelle M, Rivzi H, Long N, Plodkowaki AJ, Arbour KC, Chaft JE, Rouche JA, Zitvogel L, Zalcman G, Albiges L, Escudier B, Routy B. 2018. Negative association of antibiotics on clinical activity of immune checkpoint inhibitors in patients with advanced renal cell and non-small-cell lung cancer. Annals of Oncology. 29(6): 1437–1444

Dzutsev A, Badger JH, Perez-Chanona E, Roy S, Salcedo R, Smith CK, Trinchieri G. 2017. Microbes and Cancer. Annual Review of Immunology. 35:199–228.

Dzutsev A, Goldszmid RS, Viaud S, Zitvogel L, Trinchieri G. 2015. The role of the microbiota in inflammation, carcinogenesis, and cancer therapy. European Journal of Immunolody. 45(1):17–31.

Falk PG, Syder AJ, Guruge JL, Kischner D, Blaser MJ, Gordon JI.2000. Theoretical and experimental approaches for studying factors defining the *Helicobacter pylori*–host relationship. Trends in Microbiology.8(7): 321–329.

Farrokhi AS, Darabi N, Yousefi B, Askandar RH, Shariati M, Eslami M. 2019. Is it true that gut microbiota is considered as panacea in cancer therapy? Journal of Cellular Physiology. 1-10.

Fessler JL, Gajewski TF. 2017. The Microbiota: A New Variable Impacting Cancer Treatment Outcomes. Clinical Cancer Research. 23: 3229–3231.

Fong FLY, Kirjavainen PV, El-Nezami H. 2016. Immunomodulation of Lactobacillus rhamnosus GG (LGG)-derived soluble factors on antigen-presenting cells of healthy blood donors. Scientific Reports. 6: 22845.

Fragiadakis GK, Smits SA, Sonnenburg ED, Van-Treuren W, Reid G, Knight R, Manjurano A, Changalucha J, Dominguez-Bello MG, Leach J, Sonnenburg JL. 2019. Links between environment, diet, and the hunter-gatherer microbiome. Gut Microbes. 10(2): 216-227.

Fulbright LE, Ellermann M, Arthur JC. 2013. The microbiome and the hallmarks of cancer. Plos Pathogens. 13(9): e1006480.

Gagnaire A, Nadel B, Raoult D, Neefjes J, Gorvel JP. 2017. Collateral damage: Insights into bacterial mechanisms that predispose host cells to cancer. Nature Reviews Microbiology. 15: 109–128.

Glozak MA, Seto E. 2007. Histone deacetylases and cancer. Oncogene. 26: 5420–5432.

Gomez A, Petrzelkova KJ, Burns MB, Yeoman CJ, Amato KR, Vlckova K, Modry D, Todd A, Robimson ACJ, Remis MJ, Torralba MG, Morton E, Umana JD, Carbonero F, Gaskins HR, Nelson KE, Wilson BA, Stumpf RM, Blekhman R. 2016. Gut microbiome of coexisting baaka pygmies and bantu reflects gradients of traditional subsistence patterns. Cell Reports. 14(9): 2142-2153.

Helmink BA, Khan MAW, Hermann A, Gopalakrishnan V, Wargo J. 2019. The microbiome, cancer and cancer therapy. Nature Medicine. 25(3): 377–388

Hibberd AA, Lyra A, Ouwehand AC, Rolny P, Lindegren H, Cedgard L, Wettergren Y. 2017. Intestinal microbiota is altered in patients with colon cancer and modifed by probiotic intervention. BMJ Open GastroenterolOGY. 4(1): e000145.

IARC. 1997. Infections with Epstein-Barr virus and kaposi's sarcoma herpesvirus/ human herpesvirus. IARC monograghs on the evaluation of carcinogenic risk to humans. Lyon, France.

Iida N, Dzutsev A, Stewart CA, Smith L, Bouladoux N, Weingarten RA, Molina DA, Salcedo R, Back T, Cramer S, Dai RM, Kiu H, Cardone M, Naik S, Patri AK, Wang E, Marincola FM, Frank KM, Belkaid Y, Trinchieri G, Goldszmid RS. 2013. Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. Science. 342 (6161): 967–970.

Indiani C, Rizzardi KF, Castelo PM, Ferraz LFC, Darrieux M, Parisotto TM. 2018. Childhood obesity and firmicutes/bacteroidetes ratio in the gut microbiota: a systematic review. Childhood Obesity. 14: 501-509.

Jan G, Belzacq AS, Haouzi D, Rouault A, Metivier D, Kroemer G, Brenner C. 2002. Propionibacteria induce apoptosis of colorectal carcinoma cells via short-chain fatty acids acting on mitochondria. Cell Death Differentiation. 9: 179–188.

Jéquier E. 2002. Leptin signaling, adiposity, and energy balance. Annals of the New York Academyof Sciences. 967:379–88.

Jha AR, Davenport ER, Gautam Y, Bhandari D, Tandukar S, Ng KM, Fragiadakis GK, Holmes S, Gautam GP, Leach J, Sherchand JB, Bustamante CD, Sonnenburg JL. 2018. Gut microbiome transition across a lifestyle gradient in Himalaya. PLoS Biology. 16: e2005396.

Kew MC. 2011. Hepatitis B virus x protein in the pathogenesis of hepatitis B virus-induced hepatocellular carcinoma. Journal of Gastroenterology and Hepatology. 26(1): 144-152.

Khailova L, Baird CH, Rush AA, Barnes C, Wischmeyer PE. 2017. Lactobacillus rhamnosus GG treatment improves intestinal permeability and modulates

inflammatory response and homeostasis of spleen and colon in experimental model of *Pseudomonas aeruginosa* pneumonia. Clinical Nutrition. 36(6): 1549–1557.

Kolaček S, Hojsak I, Roberto BC, Alfredo G, Flavia I, Rok O, Bruno P, Raanan S, Hania S, Yvan V, Johannes VG, Zvi W. 2017. Commercial probiotic products: a call for improved quality control. A position paper by the ESPGHAN working group for probiotics and prebiotics. Journal of Pediatic Gastroenterology and Nutrition. 65(1): 117–124

Korpela K, Salonen A, Virta LJ, Kekkonen RA, Rorslund K, Bork P, De-Vos WM. 2016. Intestinal microbiome is related to lifetime antibiotic use in Finnish pre-school children. Nature Communications. 7: 10410.

Kostic AD, Chun E, Robertson L, Glickman JN, Gallini CA, Michaud M, Clancy TE,

Chung DC, Lochhead P, Hold GL, El-Omar EM, Brenner D, Fuchs CS, Meyerson M, Garrett WS. 2013. *Fusobacterium nucleatum* potentiates intestinal tumorigenesis and modulates the tumor-immune microenvironment. Cell Host Microbe. 14(2):207–15.

Kuper H, Adami OH, Trichopoulos D.2000a. Infections as a major preventable cause of human cancer. Journal of Internal Medicine. 248(3): 171–183.

Kuper H, Tzonou A, Kaklamani E, Hadziyannis S, Tasopoulos N, Lagiou P, Trichopoulos D, Stuver S. 2000b. Hepatitis B and C viruses in the etiology of hepatocellular carcinoma; a study in Greece using thirdgeneration assays. Cancer Causes Control. 11(2): 171–175.

Lax AJ, Thomas W. 2006. How bacteria could cause cancer: one step at a time. Trends in Microbiology.10(6): 293-299.

Lee CS, Ryan EJ, Doherty GA. 2014. Gastro-intestinal toxicity of chemotherapeutics in colorectal cancer: The role of inflammation. World Journal of Gastroenterology. 20(14): 3751–3761.

Lehouritis P, Cummins J, Stanton M, Murphy CT, McCarthy FO, Reid G, Urbaniak C, Byrne WL, Tangney M. 2015. Local bacteria affect the efficacy of chemotherapeutic drugs. Scientific Reports. 5:14554.

Ley RE, Turnbaugh PJ, KleinS, Gordon JI. 2006. Microbial ecology: human gut microbes associated with obesity. Nature. 444(7122):1022–3.

Liu H, Li J, Guo X, Liang Y, Wang W. 2018. Yeast culture dietary supplementation modulates gut microbiota, growth and biochemical parameters of grass carp. Microbial Biotechnology. 11(3): 551-565.

Magnusson PKE, Enroth H, Eriksson I, Held M, Nyren O, Engstrand L, Hansson LE, Gyllensten UB. 2001. Gastric cancer and human leukocyte antigen: distinct *DQ* and *DR* alleles are associated with development of gastric cancer and infection by *Helicobacterpylori*. Cancer Research.61(6): 2684–2689.

Mahowald MA, Rey FE, SeedorfH, Turnbaugh RJ, Fulton RS, Wollam A, Shah N, Wang C, Magrini V, Wilson RK, Cantarel BL, Coutinho PM, Henrissat B, Crock LW, Russel A, Verberkmoes NC, Hettich RL, Gordon JI. 2009. Characterizing a model human gut microbiota composed of members of its two dominant bacterial phyla. PNAS 106(14): 5859-5864.

Medina-Vera I, Sanchez-Tapia M, Noriega-López L, Granados-Portillo O, Guevara-Cruz M, Flores-lopez A, Avila-Nava A, Fernandez ML, Tovar AR, Torres N. 2019. A dietary intervention with functional foods reduces metabolic endotoxaemia and attenuates biochemical

abnormalities by modifying faecal microbiota in people with type 2 diabetes. Diabetes & Metabolism. 45(2): 122-131.

Minemura M, Shimizu Y. 2015. Gut microbiota and liver diseases. World Journal of Gastroenterology. 21(6):1691-1702.

Mira-Pascual L, Cabrera-Rubio R, Ocon S, Costales P, Parra A, Suarez A, Moris F, Rodrigo L, Mira A, Collado MC. 2015. Microbial mucosal colonic shifts associated with the development of colorectal cancer reveal the presence of different bacterial and archaeal biomarkers. Journal of Gastroenterology. 50(2):167–79.

Mohajeri MH, Brummer RJM, Rastall RA, Weersma RK, Harmsen HJM, Faas M, Eggersdorfer M. 2018. The role of the microbiome for human health: From basic science to clinical applications. European Journal of Nutrition. 57: 1–14.

Montassier E, Gastinne T, Vangay P, Al-Ghalith GA, Des Varannes SB, Moreau P, Potel G, De La Cochetiere MF, Batard E, Knights D. 2015. Chemotherapy-driven dysbiosis in the intestinal microbiome. Alimentary Pharmacology and Therapeutics. 42(5): 515–528.

Morovic W, Hibberd AA, Zabel B, Barrangou R, Stahl B. 2016. Genotyping by PCR and high-throughput sequencing of commercial probiotic products reveals composition biases. Frontiers in Microbiology. 7: 1747.

Morton ER, Lynch J, Froment A, Lafosse S, Heyer E, Przeworski M, Blekhman R, Segurel L. 2015. Variation in rural African gut microbiota is strongly correlated with colonization by entamoeba and subsistence. PLoS Genetics. 11(11): e1005658.

Nayak RR, Turnbaugh PJ. 2016. Mirror, mirror on the wall: Which microbiomes will help heal them all? BMC Medicine 14: 72

Nouri Z, Karami F, Neyazi N, Modarressi MH, Karimi R, Khorramizadeh MR, Taheri B, Motevaseli E.2016. Dual anti-metastatic and anti-proliferative activity assessment of two probiotics on HeLa and HT-29 cell lines. Cell Journal. 18(2): 127–134.

Orlandi E, Iacovellib NA, Tombolinic V, Rancatid T, Polimeni A, De Cecco L, Valdagni R, De Felice F. 2019. Potential role of microbiome in oncogenesis, outcome prediction and therapeutic targeting for head and neck cancer. Oral Oncology. 99: 104453.

Orlando A, Linsalata M, Russo F. 2016. Antiproliferative effects on colon adenocarcinoma cells induced by co-administration of vitamin K1 and Lactobacillus rhamnosus GG. International Journal of Oncology. 48(6): 2629–2638.

Pao CC, Yao DS, Lin CY, Lee SC, Ho Ting Su HT, Lin SC. 1996. Genomic aberrations of human papillomavirus recovered from cervical cancers. Biochemical and Biophysical Research Communications. 222: 116–120.

Perez-Chanona E, Trinchieri G. 2016. The role of microbiota in cancer therapy. Current Opinion in Immunology. 39:75–81.

Pisani P, Parkin MD, Munoz N, Ferlay J. 1997. Cancer and infection: estimates of the attributable fraction in 1990. Cancer Epidemiology Blomarkers and Prevention. 6(6): 387-400.

Podolsky SH. 2012. Metchnikof and the microbiome. Lancet. 380: 1810–1811.

Prieto I, Hidalgo M, Segarra AB, Martinez-Rodriguez AM, Cobo A, Ramirez M, Abriouel H, Galvez A, Martinez-Canamero M. 2018. Influence of a diet enriched

with virgin olive oil or butter on mouse gut microbiota and its correlation to physiological and biochemical parameters related to metabolic syndrome. Plos One. 13(1): e0190368.

Raza MH, Gul K, Arshad A, Riaz N, Waheed U, Rauf A, Aldakheel F, Alduraywish S, Rehman MU, Abdullah M, Arshad M. 2018. Microbiota in cancer development and treatment. Journal of Cancer Research and Clinical Oncology.145:49–63.

Riehl TE, Alvarado D, Ee X, Zuckerman A, Foster L, Kapoor V, Thotala D, Ciorba MA, Stenson WF. 2018. Lactobacillus rhamnosus GG protects the intestinal epithelium from radiation injury through release of lipoteichoic acid, macrophage activation and the migration of mesenchymal stem cells. Gut. 68(6): 1003-1013.

Robinson WS. 1994. Molecular events in the pathogenesis of hepadnavirus-associated hepatocellular carcinoma. Annual Review Medicine. 45: 297–301.

Roy S, Trinchieri G. 2017. Microbiota: a key orchestrator of cancer therapy. Nature Reviews Cancer. 17(5): 271-285.

Schéle E, Grahnemo L, Anesten F, Hallen A, Backhed F, Jansson JO. 2013. The gut microbiota reduces leptin sensitivity and the expression of the obesity-suppressing neuropeptides proglucagon (Gcg) and brain-derived neurotrophic factor (Bdnf) in the central nervous system. Endocrinology. 154(10):3643–51.

Schulfer AF, Battaglia T, Alvarez Y, Bijnens L, Ruiz VE, Ho M, Robinson S, Ward T, Cox LM, Rogers AB, Knights D, Sartor RB, Blaser MJ. 2018. Intergenerational transfer of antibiotic-perturbed microbiota enhances colitis in susceptible mice. Nature Microbiology. 3: 234-242.

Sears CL, Garrett WS. 2014. Microbes, microbiota, and colon cancer. Cell Host Microbe.15:317–28.

Shen J, Obin MS, Zhao L. 2013. The gut microbiota, obesity and insulin resistance. Molecular Aspects of Medicine. 34:39–58

Shirota Y, Shirota H, Klinman DM. 2012. Intratumoral Injection of CpG Oligonucleotides induces the differentiation and reduces the immunosuppressive activity of myeloid-derived suppressor cells. Journal of Immunology. 188: 1592–1599.

Sivan A, Corrales L, Hubert N, Williams JB, Aquino-Michaels K, Earley ZM, Benyamin FW, Lei YM, Jabri B, Alegre ML, Chang EB, Gajewski T. 2015. Commensal Bifidobacterium promotes antitumor immunity and facilitates anti-PD-L1 efficacy. Science. 350(6264):1084-1089.

Smith PA. 2015. Brain, meet gut: neuroscientists are probling the connections between intestinal microbes and brain development. Nature. 526(7573): 312-313.

Sonnenburg ED, Sonnenburg JL. 2014. Starving our microbial self: the deleterious consequences of a diet deficient in microbiota-accessible carbohydrates. Cell Metabolism. 20(5): 779-786.

Soyucak A, Zafer Yurt MN, Altunbas O, Ozalp VC, Sudagıdan M. 2020. Metagenomic and chemical analysis of tarhana during traditional fermentation process. Food Bioscience. 39: 100824.

Tomasetti C, Vogelstein B. 2015. Variation in cancer risk among tissues can be explained by the number of stem cell divisions. Science. 347(6217): 78–81.

Vaccarella S, Franceschi S, Herrero R, Munoz N, Snijders PJF, Clifford GM, Smith JS, Lazcano-Ponce E, Sukvirach S, Shin HR, Sanjose SD, Molano M, Matos E, Ferreccio C, Anh PTH, Thomas JO, Meijer CJLM, IARC. 2006. Sexual behavior, condom use, and human

papillomavirus: pooled analysis of the IARC human papillomavirus prevalence surveys. Cancer Epidemiology Biomarkers and Prevention. 15(2): 326-333.

Viaud S, Saccheri F, Mignot G, Yamazaki T, Daillere R, Hannani D, Enot DP, Pfirschke C, Engblom C, Pittet MJ, Schlitzer A, Ginhoux F, Apetoh L, Chachaty E, Woerther PL, Eberl G, Berard M, Ecobichon C, Clermont D, Bizet C, Gaboriau-Routhiau V, Cerf-Bensussan N, Opolon P, Yessaad N, Vivier E, Ryffel B, Elson CO, Dore J, Kroemer G, Lepage P, Boneca IG, Ghiringhelli F, Zitvogel L. 2013. The intestinal microbiota modulates the anticancer immune effects of cyclophosphamide, Science.342(6161): 971–976.

Wang Y, Liu L, Moore DJ, Shen X, Peek RM, Acra SA, Li H, Ren X, Polk DB, Yan F. 2017. An LGG-derived protein promotes IgA production through upregulation of APRIL expression in intestinal epithelial cells. Mucosal Immunology. 10: 373–384.

Wei W, Sun W, Yu S, Yang Y, Ai L. 2016. Butyrate production from high-fiber diet protects against lymphoma tumor. Leukemia Lymphoma. 57: 2401–2408.

Wirth U, Garzetti D, Jochum LM, Spriewald S, Kühn F, Ilmer M, Lee SML, Niess H, Bazhin AV, Andrassy J, Werner J, Stecher B, Schiergens TS. 2020. Microbiome analysis from paired mucosal and fecal samples of a colorectal cancer biobank. Cancers. 12(12): 3702.

Zhang W, Zhu YH, Yang GY, Liu X, Xia B, Hu X, Su JH, Wang JF. 2017. *LactobasillusrhamnosusGG* affects microbiota and suppresses autophagy in the intestines of pigs challenged with *Salmonella infantis*. Frontiers Microbiology. 8: 2705.

Zhao BB, Meng J, Zhang QX, Kang TT, Lu RR. 2017. Protective effect of surface layer proteins isolated from four Lactobacillus strains on hydrogen-peroxide-induced HT-29 cells oxidative stress. International Journalof Biological Macromolecules. 102: 76–83.

Zhu Y, Michelle Luo T, Jobin C, Young HA. 2011. Gut microbiota and probiotics in colon tumorigenesis. Cancer Letters. 309(2): 119–127.

Zitvogel L, Ma Y, Raoult D, Kroemer G, Gajewski TF. 2018. The microbiome in cancer immunotherapy: Diagnostic tools and therapeutic strategies. Science. 359(6382): 1366-1370.

Zmora N, Suez J, Elinav E. 2019. You are what you eat: diet, health and the gut microbiota Nature Reviews Gastroenterology Hepatology. 16(1): 35-56.